

## AMENDMENTS TO THE SPECIFICATION

Please amend Table 1 on page 10 as follows:

TABLE I

Type of electrode (w/w)	Analyte	$K_m^{app}$ ( $\mu M$ )	$I_{max}$ ( $\mu A$ )	S ( $mA/Mcm^2$ )	C (%)	DL ( $\mu M$ )
AO 87%	Histamine	279 $\pm$ 16	1.03 $\pm$ 0.02	50.57 $\pm$ 0.82	19.0	0.16
HRP 13%	Putrescine	153 $\pm$ 15	1.96 $\pm$ 0.06	175.48 $\pm$ 1.40	66.2	0.06
	H <sub>2</sub> O <sub>2</sub>	93 $\pm$ 3	1.80 $\pm$ 0.21	265.13 $\pm$ 1.65	-	-
AO 80%	Histamine	332 $\pm$ 17	<del>1.03<math>\pm</math>0.03</del> 1.34 $\pm$ 0.03	55.28 $\pm$ 0.76	16.6	0.20
HRP 20%	Putrescine	228 $\pm$ 15	3.01 $\pm$ 0.07	180.84 $\pm$ 0.95	54.7	0.07
	H <sub>2</sub> O <sub>2</sub>	112 $\pm$ 8	2.07 $\pm$ 0.06	330.23 $\pm$ 1.02	-	-
AO 67%	Histamine	370 $\pm$ 22	1.30 $\pm$ 0.03	48.13 $\pm$ 0.14	14.7	0.25
HRP 33%	Putrescine	240 $\pm$ 15	3.10 $\pm$ 0.01	176.94 $\pm$ 0.87	54.2	0.07
	H <sub>2</sub> O <sub>2</sub>	153 $\pm$ 6	3.64 $\pm$ 0.04	325.90 $\pm$ 0.56	-	-
AO 50%	Histamine	437 $\pm$ 43	1.22 $\pm$ 0.04	38.24 $\pm$ 1.42	12.7	0.33
HRP 50%	Putrescine	268 $\pm$ 23	3.05 $\pm$ 0.10	155.90 $\pm$ 1.26	52.0	0.08
	H <sub>2</sub> O <sub>2</sub>	175 $\pm$ 8	3.83 $\pm$ 0.05	299.80 $\pm$ 0.65	-	-
AO 40%	Histamine	441 $\pm$ 23	1.16 $\pm$ 0.02	36.03 $\pm$ 0.75	10.9	0.34
HRP 60%	Putrescine	276 $\pm$ 22	3.69 $\pm$ 0.06	183.14 $\pm$ 1.11	55.7	0.13
	H <sub>2</sub> O <sub>2</sub>	206 $\pm$ 3	4.94 $\pm$ 0.03	328.50 $\pm$ 0.22	-	-
AO 33%	Histamine	479 $\pm$ 41	1.37 $\pm$ 0.10	39.18 $\pm$ 1.54	12.2	0.41
HRP 67%	Putrescine	287 $\pm$ 12	3.84 $\pm$ 0.06	183.28 $\pm$ 0.61	57.0	0.08
	H <sub>2</sub> O <sub>2</sub>	211 $\pm$ 18	4.95 $\pm$ 0.15	321.36 $\pm$ 1.24	-	-

*Please amend the paragraph beginning at page 12, lines 12 as follows:*

The influence of various components of the redox hydrogel on the biosensor characteristics is shown in Table IV. The increasing  $K_m^{app}$  in the presence of both PVI<sub>13</sub>-dmeOs and PEGDGE demonstrated that the diffusion of the substrate was limited. This was because of the barrier formed by the mediator and/or cross-linking agent (rigidity of the redox hydrogel) on the surface of the electrode, which also resulted in an increased linear dynamic range. On the other hand, in the presence of crosslinked redox polycationic mediator (PVI<sub>13</sub>-dmeOs), the  $I_{max}$  value was 100% increased suggesting that the final reduction step of the topa cofactor on the electrode surface is the rate-limiting step in the absence of the ~~methyldiator~~ mediator